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(FILE 'HOME' ENTERED AT 08:07:23 ON 06 NOV 2007)  
FILE 'REGISTRY' ENTERED AT 08:07:47 ON 06 NOV 2007

L1 STRUCTURE UPLOADED

L2 1 S L1

L3 49 S L1 FULL

E ALEXA/CN

L4 1 S E15

L5 2 S E73-74

L6 1 S L3 AND L4

SEL NAME L6

FILE 'CA' ENTERED AT 08:16:44 ON 06 NOV 2007

L7 137 S L6 OR E1-2

L8 16 S ALEXA594 OR ALEXA FLUOR594 OR ALEXAFLUOR 594 OR ALEXAFLUOR594

L9 142 S L7-8

L10 5 S L8 AND PY<2003

L11 50 S L9 AND PATENT/DT

L12 25 S L11 AND PY<2005

FILE 'BIOSIS' ENTERED AT 08:23:34 ON 06 NOV 2007

L13 5 S L10

FILE 'MEDLINE' ENTERED AT 08:23:55 ON 06 NOV 2007

L14 5 S L10

FILE 'CA, BIOSIS, MEDLINE' ENTERED AT 08:24:37 ON 06 NOV 2007

L15 33 DUP REM L10 L12 L13 L14 (7 DUPLICATES REMOVED)

=> d bib,ab,kwic l15 1-33

L15 ANSWER 19 OF 33 CA COPYRIGHT 2007 ACS on STN

AN 138:283610 CA

TI New fluorescent labeling technologies for ultrasensitive cytochemical and histochemical imaging

AU Johnson, Iain

CS Molecular Probes, Inc, USA

SO Microscopy Today (2002), 10(4), 12, 14

AB Mol. Probes, Inc. introduced two novel fluorescent labeling technologies for ultrasensitive cytochem. and histochem. imaging. One of these is the Alexa Fluor series of dyes which optimizes spectroscopic and phys. properties that enable the prepn. of bioconjugates with consistently strong and photostable fluorescence output. Functionally important characteristics of Alexa Fluor dyes include strong absorption at the output wavelengths of common excitation sources, resistance to photobleaching and self-quenching, and water soly. to facilitate coupling reactions with proteins and other biomols. The well-differentiated spectra of the Alexa Fluor dyes provide many options for multicolor labeling and mol. proximity detection via fluorescence resonance energy transfer. The second novel technol. is the Zenon immunolabeling technol. which provides a rapid, convenient and completely flexible technique for coupling dyes to antibodies. Zenon technol. is based on dye- or enzyme-labeled Fab fragments of secondary antibodies directed against the Fc regions of primary antibodies.

L15 ANSWER 29 OF 33 CA COPYRIGHT 2007 ACS on STN

AN 132:233984 CA

TI Energy transfer compositions comprising phycobiliproteins  
 IN Haugland, Richard P.; Haugland, Rosaria P.  
 PA Molecular Probes, Inc., USA  
 SO PCT Int. Appl., 64 pp.  
 PI WO 2000017650 A1 20000330 WO 1999-US22193 19990923  
 WO 9915517 A1 19990401 WO 1998-US19921 19980923  
 PRAI WO 1998-US19921 W 19980923  
 US 1998-209045 A 19981209  
 US 1997-935963 A 19970923  
 WO 1999-US22193 W 19990923  
 AB Energy transfer compns. comprising one or more fluorescent dyes and a fluorescent protein are described, in particular where the fluorescent dye is a sulfonated dye and the fluorescent protein is a phycobiliprotein. The energy transfer compns. of the invention may further comprise addnl. fluorescent dyes or fluorescent proteins that act as intermediate energy transfer dyes or ultimate emitter dyes. The energy transfer compns. of the invention may also be substituted by chem. reactive functional groups, or covalently bound conjugated substances. The compns. of the invention possess utility as detection reagents and as fluorescent tracers in a wide variety of applications, including biol. applications.

L15 ANSWER 31 OF 33 CA COPYRIGHT 2007 ACS on STN  
 AN 134:97365 CA  
 TI Light-Induced Conformational Changes of Rhodopsin Probed by Fluorescent Alexa594 Immobilized on the Cytoplasmic Surface  
 AU Imamoto, Yasushi; Kataoka, Mikio; Tokunaga, Fumio; Palczewski, Krzysztof  
 CS Graduate School of Materials Science, Nara Institute of Science and Technology, Ikoma Nara, 630-0101, Japan  
 SO Biochemistry (2000), 39(49), 15225-15233  
 AB A novel fluorescence method has been developed for detecting the light-induced conformational changes of rhodopsin and for monitoring the interaction between photolyzed rhodopsin and G-protein or arrestin. Rhodopsin in native membranes was selectively modified with fluorescent Alexa594-maleimide at the Cys316 position, with a large excess of the reagent Cys140 that was also derivatized. Modification with Alexa594 allowed the monitoring of fluorescence changes at a red excitation light wavelength of 605 nm, thus avoiding significant rhodopsin bleaching. Upon absorption of a photon by rhodopsin, the fluorescence intensity increased as much as 20% at acidic pH with an apparent pKa of ~6.8 at 4°, and was sensitive to the presence of hydroxylamine. These findings indicated that the increase in fluorescence is specific for metarhodopsin II. In the presence of transducin, a significant increase in fluorescence was obsd. This increase of fluorescence emission intensity was reduced by addn. of GTP, in agreement with the fact that transducin enhances the formation of metarhodopsin II. Under conditions that favored the formation of a metarhodopsin II-Alexa594 complex, transducin slightly decreased the fluorescence. In the presence of arrestin, under conditions that favored the formation of metarhodopsin I or II, a phosphorylated, photolyzed rhodopsin-Alexa594 complex only slightly decreased the fluorescence intensity, suggesting that the cytoplasmic surface structure of metarhodopsin II is different in the complex with arrestin and transducin. These results demonstrate the

application of Alexa594-modified rhodopsin (Alexa594-rhodopsin) to continuously monitor the conformational changes in rhodopsin during light-induced transformations and its interactions with other proteins.

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STN INTERNATIONAL LOGOFF AT 08:25:24 ON 06 NOV 2007